

**7-Methoxy-2,3-dimethylbenzofuran-5-ol,
a New Antioxidant from *Malbranchea
cinnamomea* HKI 0286**

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In the course of our continuing screening for inhibitors of xanthine oxidase and peroxidase as key enzymes of purine metabolism and oxygen radical generation^{1,2)} we found the culture extract of the fungus *Malbranchea cinnamomea* HKI 0286 to contain an inhibitory principle. Here we report the isolation and properties of 7-methoxy-2,3-dimethylbenzofuran-5-ol (**1**; Figure 1) as a new natural antioxidant.

The fungal strain *Malbranchea cinnamomea* HKI 0286 from the strain collection of Hans-Knöll-Institute for Natural Products Research (Jena, Germany) was cultivated in 1 liter Erlenmeyer flasks containing 250 ml of a medium composed as follows (g/liter): dextrose 10, malt extract 20, soybean meal 2, yeast extract 1, KH₂PO₄ 1, and MgSO₄×7H₂O 0.5, pH 6.0. The cultivation was done on

rotary shakers (110 r.p.m.) for 14 days at 35°C.

After 14 days the culture broth (14 liters) was extracted three times with ethyl acetate, and the combined extracts were dried and evaporated to yield 2.1 g of a crude product. Isolation of **1** was carried out by several subsequent chromatographic steps such as column chromatography on silica gel 60 (Merck, 0.063~0.1 mm, CHCl₃/MeOH, 95 : 5, v/v) and preparative TLC on silica gel aluminium sheets (Merck) using the same eluent. Compounds on the chromatograms were detected by their staining behaviour in the presence of 1% vanillin in conc. H₂SO₄. Finally 25 mg of **1** with R_f 0.9 on TLC staining bluish were obtained as a solid compound. After some days standing at ambient

Fig. 1. Structure of 7-methoxy-2,3-dimethylbenzofuran-5-ol (**1**) from *Malbranchea cinnamomea* HKI 0286.

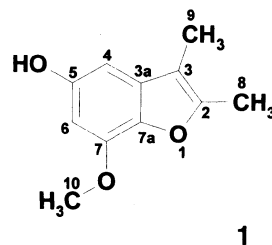


Table 1. Physico-chemical properties of **1**.

	1
Appearance	Semicrystalline solid
M.P.	108 – 110 °C
Molecular weight (M ⁺ ; HREI-MS)	192.0780 (calcd. 192.0786)
Chemical formula	C ₁₁ H ₁₂ O ₃
UV λ ^{MeOH} _{max} ; nm (ε)	215 (6.300), 312 (2.835)
IR (λ _{max} ; cm ⁻¹)	751, 836, 966, 1049, 1118, 1126, 1188, 1216, 1248, 1292, 1364, 1399, 1439, 1493, 1555, 1599, 1633, 2920, 3428
R _f on TLC Silica gel (CHCl ₃ /MeOH, 95:5)	0.9

† Deceased February 14, 2003.

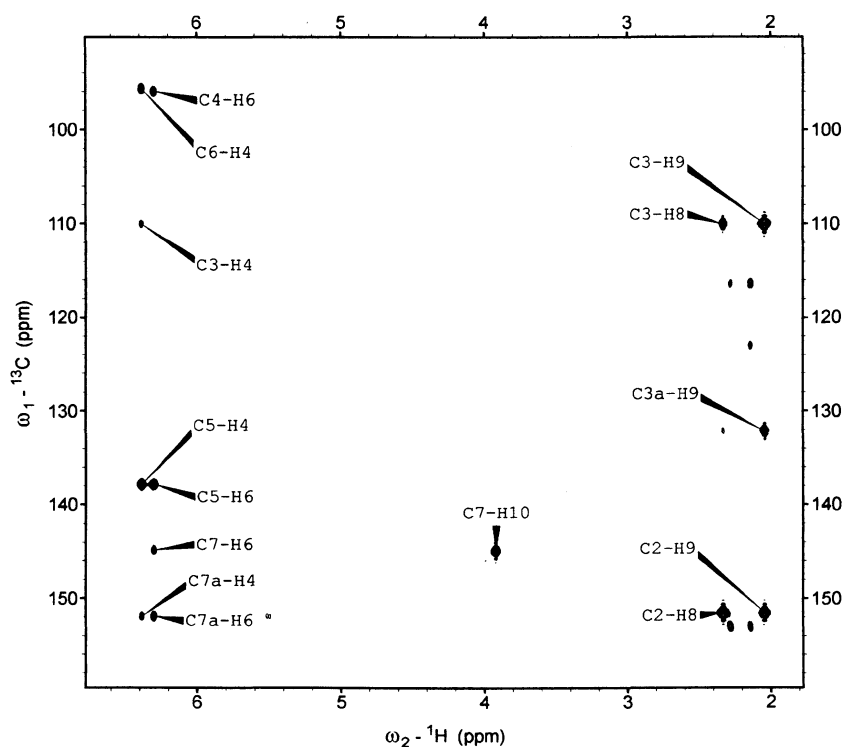
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temperature the spot of **1** on the chromatogram turned reddish due to autoxidation and formation of quinoid products. The physico-chemical properties of **1** are shown in Table 1. The structure of **1** as shown in Figure 1 was assigned on the basis of optical spectroscopy (UV, FTIR, polarimetry), mass spectrometry (ESI-MS, HREI-MS) and 1D/2D NMR spectroscopy (^1H , ^{13}C , DEPT, ^1H , ^1H -COSY, HMQC, HMBC). **1** was optically inactive. The FTIR spectrum suggested the occurrence of aromatic structures (1555 , 1599 , 1633 cm^{-1}) and a hydroxyl group (3428 cm^{-1}). The ESI-MS showed m/z 193.2 ($[\text{M}+\text{H}]^+$), and in the HREI-MS m/z 192.0780 (M^+ ; 100 %, calcd. 192.0786 for $\text{C}_{11}\text{H}_{12}\text{O}_3$) was visible. The ^{13}C NMR spectrum (CDCl_3) displayed 11 carbons which were assigned on the basis of the DEPT spectrum as two methyls (11.8 ppm (C-8), 8.0 ppm (C-9)), one methoxyl (56.0 ppm (C-10)), four oxygenated olefinic or aromatic carbons (151.5 ppm (C-2), 151.8 ppm (C-7a), 144.8 ppm (C-7), 137.8 ppm (C-5)), two protonated aromatic carbons (95.7 ppm (C-6), 97.0 ppm (C-4)) and two additional quaternary carbons at 132.1 ppm (C-3a) and 110.0 ppm (C-3). In the ^1H NMR spectrum 12 protons appeared at 2.33 ppm (H-8, s, 3H), 2.04 ppm (H-9, s, 3H), 3.92 (H-10,

s, 3H), 6.39 ppm (H-4, d, 1.8 Hz, 1H), 6.30 ppm (H-6, d, 1.8 Hz, 1H), and one broad proton signal at 4.92 ppm (OH-5, br).

The COSY spectrum and $^4J_{\text{H-4,H-6}}=1.8\text{ Hz}$ indicated the presence of meta-coupled protons. Conclusive evidence for the structure of **1** was furnished by C,H long-range coupled NMR spectra (HMBC, Fig. 2) showing a series of instructive $^2J_{\text{C,H}}$ and $^3J_{\text{C,H}}$ couplings which enabled us together with the three couplings from the NOESY spectrum doubtlessly the assignment of carbon and proton signals (Table 2). Especially the NOE between H-9 (methyl) and H-4 gives us the evidence for the position of C-4. The couplings from the HMBC and the NOESY spectrum are shown in Fig. 3. 7-Methoxy-2,3-dimethylbenzofuran-5-ol (**1**) thus appears as a new representative of the benzofuran family of natural products. It displayed strong inhibition of xanthine oxidase¹⁾ with IC_{50} $9.8\ \mu\text{M}$ and horse radish peroxidase with IC_{50} $7.5\ \mu\text{M}$. Moreover, **1** inhibited the polymorphonuclear lymphocyte reaction²⁾ (preliminary results). The inhibitory properties of **1** may be due to the presence of a cryptic hydroquinone structure similar as occurring in the tocopherol analogue Trolox^{®3)} which can be oxidized easily to the quinone.

Fig. 2. HMBC spectrum of **1**.



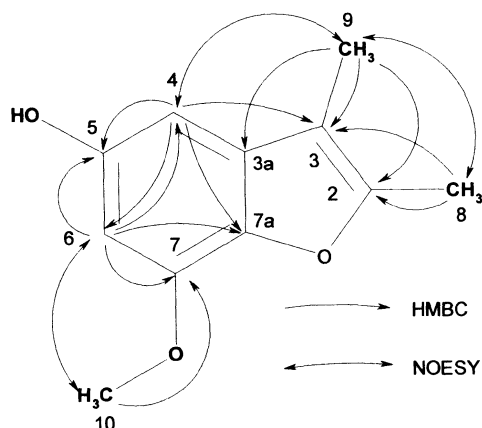
The peaks are marked with the corresponding atom numbers. Unmarked peaks are due to minor impurities.

Table 2. Assignment of ^1H and ^{13}C NMR spectra of **1** (500 MHz; in CDCl_3 ; chemical shifts (δ) in ppm; coupling constants in Hz).

Position	1	
	δ_{C}	δ_{H}
2	151.5 (s)	-
3	110.0 (s)	-
3a	132.1 (s)	-
4	97.0 (d)	6.39 d, 1.8
5	137.8 (s)	4.92 br (OH)
6	95.7 (d)	6.30 d, 1.8
7	144.8 (s)	-
7a	151.8 (s)	-
8	11.8 (q)	2.33 (s)
9	8.0 (q)	2.04 (s)
10	56.0 (q)	3.92 (s)

Abbreviations: s=singlet, d: doublet, br: broad, q=quartet.

Fig. 3. Long range couplings from HMBC and NOESY spectra.



Experimental

UV spectra were recorded on a Specord double-beam 2000 spectrometer (Analytik Jena, Germany), FTIR on a Mattson FTIR spectrometer satellite equipped with an ATR device (Mattson, Chicago, USA), ESI mass spectra on a triple quadrupole instrument Quattro (VG Biotech, Altrincham, England), HREI-MS on a Finnigan MAT 95 XL (Finnigan, Bremen, Germany), and NMR spectra on a 500 MHz Bruker Avance DRX 500 spectrometer (Bruker, Karlsruhe, Germany). Measurements of xanthine oxidase¹⁾ activity and polymorphonuclear lymphocyte reaction²⁾ were done as was described in the literature. Horse radish peroxidase was measured by formation of 5-amino-2,3-

dihydro-1,4-phthalazinedione (luminol) radicals and endoperoxides in presence of H_2O_2 . Their decomposition yielded 3-aminophthalate dianion which was excitable to emit visible light. This chemiluminescence was measured as relative light units. The addition of **1** to the reaction mixture of a single assay in a total volume of 250 μl per well of a 96 well microplate (50 μl luminol (0.4 mg/ml PBS buffer), 50 μl horse radish peroxidase (Sigma; 0.066 U/ml, 1: 0.1~10 μg in 50 μl PBS buffer, 50 μl H_2O_2 (30%) and PBS buffer ad 250 μl) resulted in continuous suppression of light emission until the antioxidant (**1**) was consumed. Chemiluminescence was measured by a labsystem Luminoskan RS instrument.

Acknowledgement

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